



The Brazilian Journal of INFECTIOUS DISEASES

www.elsevier.com/locate/bjid



Original Article

Investigation of etiology of community-acquired pneumonia in hospitalized patients in a tertiary hospital of São Paulo City, Brazil

Daniel Joelsons, MD, PhD ^{a,*}, Cecília Salete Alencar, PhD ^b,
João Renato Rebello Pinho, MD, PhD ^{b,c}, Yeh-Li Ho, MD, PhD ^a

^a Universidade de São Paulo, Hospital das Clínicas, Faculdade de Medicina (HCFMUSP), Departamento e Divisão de Moléstias Infecciosas e Parasitárias, São Paulo, SP, Brazil

^b Universidade de São Paulo, Hospital das Clínicas, Faculdade de Medicina (HCFMUSP), Laboratório de Medicina Laboratorial – Divisão de Laboratório Central, São Paulo, SP, Brazil

^c Universidade de São Paulo, Hospital das Clínicas, Faculdade de Medicina (HCFMUSP), São Paulo, SP, Brazil

ARTICLE INFO

Article history:

Received 5 July 2023

Accepted 31 October 2023

Available online 13 November 2023

Keywords:

Community-acquired pneumonia

Severe pneumonia

Brazil

Etiology

Multiplex polymerase chain reaction

ABSTRACT

Background: Community-Acquired Pneumonia (CAP) is the primary cause of hospitalization in the United States and the third leading cause of death in Brazil. The gold standard for diagnosing the etiology of CAP includes blood culture, Gram-stained sputum, and sputum culture. However, these methods have low sensitivity. No studies investigating the etiology of CAP have been conducted in Brazil in the last 20-years, and the empirical choice of antimicrobials is mainly based on the IDSA guidelines. This is the first national study with this aim, and as a result, there's potential for the Brazilian consensus to be impacted and possibly modify its guidelines rather than adhering strictly to the IDSA's recommendations.

Methods: The aim of this study is to identify the main microorganisms implicated in CAP by employing a multiplex Polymerase Chain Reaction (mPCR) at the foremost public hospital in Brazil. All patients who were admitted to the emergency department and diagnosed with severe CAP underwent an mPCR panel using nasopharyngeal and oropharyngeal swabs, with the aim of detecting 13 bacterial and 21 viral pathogens.

Results: A total of 169 patients were enrolled in the study. The mPCR panel identified an etiological agent in 61.5% of patients, with viruses being the most common (42.01%), led by Rhinovirus, followed by Influenza and Coronavirus (non-SARS-CoV-2). Bacterial agents were identified in 34.91% of patients, with *S. pneumoniae* being the most common, followed by *H. influenzae*, *M. catarrhalis*, and *S. aureus*. Additionally, we found that the prescription for 92.3% of patients could be modified, with most changes involving de-escalation of antibiotics and antiviral therapy.

* Corresponding author.

E-mail address: daniel.joelsons@einstein.br (D. Joelsons).

<https://doi.org/10.1016/j.bjid.2023.103690>

1413-8670/© 2023 Sociedade Brasileira de Infectologia. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

Conclusion: Our study revealed different etiological causes of CAP than those suggested by the Brazilian guidelines. Using molecular diagnostic tests, we were able to optimize treatment by using fewer antibiotics.

© 2023 Sociedade Brasileira de Infectologia. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>)

Introduction

Lower respiratory infections are one of the top ten causes of death according to the World Health Organization (WHO), and Community-Acquired Pneumonia (CAP) is the main cause of hospitalization in the United States, with medical costs exceeding \$10 billion in 2011. CAP is also the third leading cause of death in Brazil.¹⁻⁴

Traditionally, clinicians have used X-Rays in combination with clinical symptoms as a standard tool to diagnose pneumonia, however, several other causes can lead to the same findings. Additionally, until recent years, the gold standard for diagnosing the etiology of pneumonia was blood culture, Gram-stained sputum, and sputum culture. However, these methods have low sensitivity, and more than 50% of patients either do not produce sputum or have sputum of poor quality for analysis.⁵⁻¹¹

The guideline of Infectious Diseases Society of America (IDSA) is commonly used to guide the diagnosis and treatment of CAP worldwide. Despite limited evidence, the guideline do not recommend using etiological diagnostic tests for CAP except in severe condition and maintain the same bacterial agents over the last decade (*S. pneumoniae*, *H. influenzae*, *M. pneumoniae*, *S. aureus*, *Legionella spp*, *C. pneumoniae*, and *M. catarrhalis*) as the main CAP pathogens. Nevertheless, a recent study conducted in the United States drew parallels between the etiological agents documented and those delineated in its respective guideline, revealing a concordance rate of under 30%, the agreement will be even less when another country, such as Brazil, uses them as a foundation for its own guidelines.¹²⁻¹⁵

The Polymerase Chain Reaction (PCR) has been shown to be more sensitive than traditional diagnostic methods for determining the etiology of CAP.^{16,17} Recent studies using this method have identified different etiologies than those suggested by the guidelines, including respiratory viruses.^{3,18}

No studies investigating the etiology of CAP have been conducted in Brazil in the last 20-years, and the empirical choice of antimicrobials is mainly based on the IDSA guidelines. The aim of this study is to use a multiplex Polymerase Chain Reaction (mPCR) panel in conjunction with traditional diagnostic methods (blood culture, Gram-stained sputum, and sputum culture) to identify the main microorganisms responsible for community-acquired pneumonia at the largest public hospital in Brazil.

Methods

Study design

From September 2017 to August 2018, we screened all patients admitted to the emergency room of Hospital das Clínicas da

Faculdade de Medicina da Universidade de Sao Paulo (ER-HC-FMUSP) who required hospitalization based on clinician decision. ER-HC-FMUSP is a teaching hospital that serves as a reference for all severe clinical and surgical cases. During this period, there were 6,205 clinical admissions. We attempted to enroll all eligible adults between Sunday and Thursday over the course of one year in order to include all seasonal pathogens. Due to laboratory operations, enrollment was limited to these days. Written informed consent was obtained from all patients or their caregivers prior to enrollment

Patients were enrolled if they were older than 18-years, had evidence of CAP defined as lung imaging (chest radiography or computed tomography), and two of the following symptoms: fever (temperature >37.8°C), cough, sputum production, shortness of breath, pleuritic chest pain, mental confusion, leukocytosis (White Blood Count [WBC] >12,000 mm³), or a suppressed WBC count (<6,000 mm³).

Patients were excluded if they had been recently hospitalized (<30 days), admitted more than 48 hours before enrollment, undergone hematopoietic stem cell transplantation, received chemotherapy in the past 30 days, received antibiotics for over 48 hours before enrollment, were postoperative, or had a clear alternative diagnosis.

Severity was evaluated using the Pneumonia Severity Index (PSI) Score.¹⁹

Antibiotic appropriateness was verified using the "Sanford Guide Antimicrobial Stewardship" book.²⁰

The primary outcome was to identify the main microorganisms responsible for community-acquired pneumonia at the largest public hospital in Brazil. The secondary outcome involves assessing diagnostic gain using the mPCR methodology compared to standard methodology and the appropriateness of the prescribed antibiotic.

Specimen collection

Blood cultures, sputum (if patients had productive cough), nasopharyngeal and oropharyngeal swabs were obtained from all patients after obtaining their consent. The results of the PCR tests were not available until the end of the study and were not used for patient care.

Laboratory testing

Blood and high-quality sputum samples were collected using standard techniques.

The mPCR panel was performed on nasopharyngeal and oropharyngeal swabs. For the first 107 patients, the Mobius Life Science's kit was used, which identified the following pathogens: Influenza A; Influenza B; Influenza C; Influenza A (H1N1); Parainfluenza viruses 1, 2, 3 and 4; Coronaviruses

NL63, 229E, OC43 and HKU1; Human metapneumoviruses A/B; Rhinovirus; Respiratory syncytial viruses A and B; Adenovirus; Enterovirus; Parechovirus; Bocavirus; *Pneumocystis jirovecii*; *Mycoplasma pneumoniae*; *Chlamydia pneumoniae*; *Streptococcus pneumoniae*; *Haemophilus influenzae*; *Haemophilus influenzae* type B; *Staphylococcus aureus*; *Moraxella catarrhalis*; *Bordetella* spp.; *Klebsiella pneumoniae*; *Legionella pneumophila*; *Legionella longbeachae* and *Salmonella* spp. However, due to the discontinuation of the Mobius Life Science's kit in the hospital laboratory, the Biomerieux's Biofire FilmArray kit was used for the last 57 patients. This kit identified: Influenza A; Influenza B; Influenza A (H1N1); Parainfluenza viruses 1, 2, 3 and 4; Coronaviruses NL63, 229E, OC43 and HKU1; Human metapneumoviruses A/B; Rhinovirus/Enterovirus; Respiratory syncytial viruses A and B; Adenovirus; *Mycoplasma pneumoniae*; *Chlamydia pneumoniae*; *Bordetella parapertussis* and *Bordetella pertussis*.

Statistical analysis and ethics

All data was stored on the Redcap platform. We used prevalence, mean, and standard deviation for parametric data, and median and interquartile range (p25%–p75%) for non-parametric data. The Shapiro-Wilk test was used to determine if the data was normally distributed, with statistical significance set at a p-value of <0.05. The analyses were performed using Stata 15.1 software.

For the *Staphylococcus aureus* bacteria results, considering its potential ability to colonize the upper airways and skin, and the fact that carriage has been reported to be highly variable between populations (4%–64%),²¹ we used a lowered threshold cycle of interquartile p25% to consider a positive test.

The study was approved by the Ethics Committee at HC-FMUSP (CAPPesq 11751), and written informed consent was obtained from all participants. The principles of the Declaration of Helsinki were followed, and the study was conducted according to good clinical practice guidelines.

Results

Out of 172 eligible patients, 169 were enrolled in the study. The mean age was 64 years and 54% of them were male. The most common comorbidity was hypertension, followed by diabetes mellitus, heart failure, and chronic obstructive pulmonary disease. About 28.6% of the patients had some immunosuppressive condition. Seven patients required intubation upon arrival at the emergency room (Table 1).

Among the 169 patients, 40% required admission to the intensive care unit, 23% progressed to respiratory support with invasive mechanical ventilation, and 13.6% required renal replacement therapy. The mean length of hospital stay was 12.8-days, and the mortality rate was 14.2% (Table 2).

The blood culture has been collected from 140 patients with only 5.7% of them with positive's results. Sputum cultures and endotracheal aspirate cultures were collected from 15 and 12 patients, respectively. The results are described in Table 3. When we analyzed the positive cases using the standard etiology investigation, we found a total of 11 cases, representing 6.52% of the total number of patients.

Table 1 – Baseline characteristics of patients with community-acquired pneumonia requiring hospitalization at the ER-HC-FMUSP.

Total	n = 169
Age, years – Median (IQR)	64 (51–76)
Gender, male – n (%)	91 (54.5)
Antibiotic administration previously to swab sampling – n (%)	134 (79.7)
Time between antibiotic administration and swab sampling, hours – Median (IQR)	13 (8–19)
Comorbidity	
Systemic arterial hypertension – n (%)	100 (59.5)
Diabetes mellitus – n (%)	55 (33.7)
Heart failure – n (%)	45 (27.1)
Chronic obstructive pulmonary disease – n (%)	40 (24.1)
Chronic kidney disease – n (%)	37 (22.2)
Dialytic – n (%)	9 (19.6)
Cerebrovascular disease – n (%)	12 (7.2)
Neoplastic disease – n (%)	10 (6.1)
Solid organ transplant – n (%)	20 (12.1)
Any Immunosuppression – n (%)	48 (28.6)
Immunosuppressive drugs – n (%)	37 (21.9)
Human immunodeficiency virus infection – n (%)	8 (4.7)
Inherited immunodeficiency syndromes – n (%)	3 (1.8)
Seasonal influenza vaccination – n (%)	61/96 (38.6)
Peripheral oxygen saturation, % – Median (IQR)	90.5 (83–95)
Mental confusion – n (%)	39 (23.2)
Pleural effusion – n (%)	37 (21.9)
Mechanical ventilation – n (%)	7 (3.9)
Pneumonia severity index	
1–3 – n (%)	35 (20.7)
4 – n (%)	71 (42.0)
5 – n (%)	63 (37.2)

The mPCR panel identified an etiological agent in 61.5% of patients, with viruses being the most common (42.01%), led by Rhinovirus, followed by Influenza and Coronavirus (non-SARS-CoV-2). Bacterial agents were identified in 34.91% of patients, with *S. pneumoniae* being the most common, followed by *H. influenzae*, *M. catarrhalis*, and *S. aureus* (Table 4).

Adopting both diagnostic methods, we found that the prescription for 92.3% of patients could be modified, with most changes involving de-escalation of antibiotics and antiviral therapy (Table 5). The primary class of antibiotics that could be subject to modification was macrolides.

Table 2 – Evolution and Mortality (total and stratified by PSI) in 169 community acquired pneumonia in the ER-HC-FMUSP.

	n (169)	%
ICU admission	68	40.2
Invasive mechanical ventilation	40	23.7
Vasopressor requirement	36	21.3
Renal replacement therapy	22	13.6
Length of stay (LOS) – days, Mean – SD	12.8	15.5
Mortality	24	14.2
PSI 1–3	2/24	8.34
PSI 4	10/24	41.67
PSI 5	12/24	50

Table 3 – Numbers and positive microorganisms identified using the standard etiology investigation in each culture sample.

Blood culture	n = 140
Negative	127
Positive	8
S. pneumoniae	1
H. influenzae	1
S. aureus	3
E. coli	1
Cryptococcus neoformans	2
Sputum culture	n = 15
Negative	10
S. pneumoniae	2
P. aeruginosa	1
M. Tuberculosis	2
Endotracheal aspirates culture	n = 12
Negative	10
S. Aureus	1
S. pneumoniae	1

Discussion

Traditional methods of etiological diagnosis for Community-Acquired Pneumonia (CAP) have been restricted to the investigation of bacterial agents and have limited sensitivity.⁵⁻⁹ These limitations have led ISDA and Brazilian consensus guidelines to not recommend etiological investigation for mild cases or in patients without comorbidities. Even for cases requiring hospitalization, obtaining respiratory samples for identification of the agent is only recommended for those with severe forms of the disease or at risk of infection with resistant bacteria.¹⁴ These recommendations are justified by

Table 4 – Pathogen detection in patients with community-acquired pneumonia molecular methods (n = 169).

Pathogens	n (169)	%
Any pathogens	104	61.54
Virus		
Any Virus	71	42.01
Rhinovirus	27	15.98
Influenza	24	14.20
Coronavirus	7	4.14
Parainfluenza virus	5	2.96
Human metapneumovirus	4	2.37
Adenovirus	4	2.37
Respiratory syncytial virus	4	2.37
Bocavirus	3	1.78
Bacteria	59	34.91
<i>Streptococcus pneumoniae</i>	27	15.98
<i>Haemophilus influenzae</i>	15	8.88
<i>Moraxella catarrhalis</i>	15	8.88
<i>Staphylococcus aureus</i>	14	8.28
<i>Klebsiella pneumoniae</i>	6	3.55
<i>Mycoplasma pneumoniae</i>	3	1.78
<i>Mycobacteria tuberculosis</i>	2	1.18
<i>Legionella spp.</i>	1	0.59
<i>Escherichia coli</i>	1	0.59
<i>Pseudomonas aeruginosa</i>	1	0.59
<i>Pneumocystis jirovecii</i>	6	3.55
<i>Cryptococcus neoformans</i>	2	1.18
Codetection of virus and bacteria	26	15.38

Table 5 – Estimated potential impact of molecular testing on treatment prescribing in patients with community-acquired pneumonia.

Potential Modification	n	%
Escalation antibiotic	3	1.78
De-escalation antibiotic	153	90.53
Escalation antiviral	17	10.06
De-escalation antiviral	20	11.83
Any change	158	92.31
No change	13	7.69

their lack of impact on clinical outcomes in addition to the cost of the tests. We observed a low frequency of collection of sputum or tracheal secretion samples, demonstrating the difficulty in obtaining these samples. In addition, despite the high frequency of blood culture collection, the positivity rate was quite low, bringing little contribution to guide the antimicrobial. These factors contribute to the recommendation of empirical use of antimicrobials in these guidelines, especially with coverage against pneumococci, as coinfection with viral and bacterial agents is common.^{12,14,22-25} This situation is further exacerbated in Brazil, where there is a lack of comprehensive knowledge regarding the primary causative agents of CAP. Consequently, the American guidelines are employed as a reference, both for determining etiological agents and for the treatment of CAP in our hospital.

However, the increasing antimicrobial resistance is becoming a global problem, including for community infections, to the point that in 2001, the World Health Organization drew attention to this scenario and outlined strategies to contain this problem, which has as its first step the rational use of antimicrobials.²⁶

The H1N1 influenza pandemic in 2009 reinforced the possibility of a viral agent causing severe acute respiratory failure and ARDS, including radiological consolidation patterns and the potential for fatal outcomes.²⁷ This scenario was further reinforced during the COVID-19 pandemic, where several authors observed that the isolated viral agent could be responsible for indistinguishable cases of bacterial pneumonia, even in patients admitted to the ICU.²⁸⁻³⁰

Our study shows that expanded etiology investigation for CAP using molecular diagnostic methods can increase pathogen detection from 6.52% to 61.54%, representing a 55% increase in positive diagnoses. With this more sensitive approach, we described a different prevalence of the principal microorganisms causing severe CAP in our country. We are the first study to investigate the etiology of severe CAP in Brazil, and we found different microorganisms compared to those described in national guidelines.²⁵ The most prevalent agent was a virus (61.54%), while bacteria were found in only 34.91% of the samples, being *S. pneumoniae* more prevalent followed by, *H. influenzae*, *M. catarrhalis*, and *S. aureus*. These results are similar to studies using molecular diagnostic tests in other countries.³

Another contribution of our study is the low prevalence of *M. pneumoniae*. In the Brazilian guideline for treating CAP, it's mentioned as the second most prevalent agent, but our findings indicate an incidence of only 1.78%. Although consensus

recommends introducing a macrolide as antibiotic therapy, we observed that this may not be necessary as a routine. Our results are consistent with recent studies worldwide that use this molecular technique.^{3,17,18}

Upon estimating the potential impact of molecular testing on the treatment of patients diagnosed CAP, we observed a significant increase in diagnostic accuracy, with the percentage rising from 6.52% to 61.5%, representing a nearly 55% rise as compared to the standard methodology.

The indiscriminate use of antibiotics in clinical practice is a concern, and in our study, we found that using a molecular diagnostic test for CAP could lead to more than 90% of treatments being changed, resulting in a significant de-escalation in antibiotic use.

A limitation of our study is that it was conducted in only one hospital and therefore cannot be extrapolated to the entire country. Further studies need to be done to confirm our hypothesis. Another limitation is that we used a nasopharyngeal swab for etiological investigation, which means that some agents may be just colonization and not the source of infection. However, as we only evaluated patients who presented clinical symptoms compatible with pneumonia and signs of severity by the PSI score, we consider that if we isolate an etiological agent that is plausible to cause infection, it should be considered as the responsible agent for the infection and not just colonization. Another limitation of our study is that patients were already using antibiotics at the time of collecting respiratory samples. This could potentially have influenced the negativity of the culture tests and PCR panel results.

In conclusion, despite our limitations, we found different etiological causes of PAC than those suggested by the Brazilian guidelines. Using molecular diagnostic tests, we were able to optimize our treatment by using fewer antibiotics.

Conflicts of interest

The authors declare no conflicts of interest.

REFERENCES

- Metlay JP, Long AC, Anzueto A, Brozek J, Crothers K, Cooley LA, et al. Diagnosis and treatment of adults with community-acquired pneumonia. An official clinical practice guideline of the American Thoracic Society and Infectious Diseases Society of America. *Am J Respir Crit Care Med.* 2019;200(7):e45–67.
- Ministério da Saúde D, SINAM. 2022 [Available from: <http://www2.datasus.gov.br/DATASUS/>].
- Jain S, Self WH, Wunderink RG, Fakhran S, Balk R, Bramley AM, et al. Community-acquired pneumonia requiring hospitalization among U.S. adults. *N Engl J Med.* 2015;373:415–27.
- Global health estimates: leading causes of death. 2022. Available from: <https://www.who.int/data/gho/data/themes/mortality-and-global-health-estimates/ghe-leading-causes-of-death>.
- Chalasanani NP, Valdecanas MAL, Copal AK, JEM Jr, Jurado RL. Clinical utility of blood cultures in adult patients with community-acquired pneumonia without defined underlying risks. *Chest.* 1995;108:932–6.
- Evans L, Rhodes A, Alhazzani W, Antonelli M, Coopersmith CM, French C, et al. Surviving sepsis campaign: international guidelines for management of sepsis and septic shock 2021. *Crit Care Med.* 2021;49(11):e1063–143.
- Campbell SG, Marrie TJ, Anstey R, Dickinson G, Ackroyd-Stolarz S. The contribution of blood cultures to the clinical management of adult patients admitted to the hospital with community-acquired pneumonia: a prospective observational study. *Chest.* 2003;123:1142–50.
- Lidman C, Burman LG, Lagergren A, Orqvist A. Limited value of routine microbiological diagnostics in patients hospitalized for community-acquired pneumonia. *Scand J Infect Dis.* 2002;34:873–9.
- Rosón B, Carratalà J, Verdaguer R, Dorca J, Manresa F, Gudiol F. Prospective study of the usefulness of sputum Gram stain in the initial approach to community-acquired pneumonia requiring hospitalization. *Clin Infect Dis.* 2000;31:869–74.
- Musher DM, Montoya R, Wanahita A. Diagnostic value of microscopic examination of Gram-stained sputum and sputum cultures in patients with bacteremic pneumococcal pneumonia. *Clin Infect Dis.* 2004;39:165–9.
- Rio-Pertuz GD, Gutiérrez JF, Triana AJ, Molineras JL, Robledo-Solano AB, Meza JL, et al. Usefulness of sputum gram stain for etiologic diagnosis in community-acquired pneumonia: a systematic review and meta-analysis. *BMC Infect Dis.* 2019;19:403.
- Mandell LA, Wunderink RG, Anzueto A, Bartlett JG, Campbell D, Dean NC, et al. Infectious Diseases Society of America/American Thoracic Society consensus guidelines on the management of community-acquired pneumonia in adults. *Clin Infect Dis.* 2007;44(Suppl 2):S27–72. Suppl 2.
- Carugati M, Aliberti S, Reyes LF, Sadud RF, Irfan M, Prat C, et al. Microbiological testing of adults hospitalized with community-acquired pneumonia: an international study. *ERJ Open Res.* 2018;4:00096–2018.
- Metlay JP, Waterer GW, Long AC, Anzueto A, Brozek J, Crothers K, et al. Diagnosis and treatment of adults with community-acquired pneumonia. *JAMA.* 2020;323(9):885–6.
- Woodhead M, Blasi F, Ewig S, Garau J, Huchon G, Ieven M, et al. Guidelines for the management of adult lower respiratory tract infections – summary. *Clin Microbiol Infect.* 2011;17(Suppl 6):1–24.
- Templeton KE, Scheltinga SA, van den Eeden WC, Graffelman AW, van den Broek PJ, Claas EC. Improved diagnosis of the etiology of community-acquired pneumonia with real-time polymerase chain reaction. *Clin Infect Dis.* 2005;41:345–51.
- Gadsby NJ, Russell CD, McHugh MP, Mark H, Conway Morris A, Laurenson IF, et al. Comprehensive molecular testing for respiratory pathogens in community-acquired pneumonia. *Clin Infect Dis.* 2016;62:817–23.
- Di Pasquale MF, Sotgiu G, Gramegna A, Radovanovic D, Terraneo S, Reyes LF, et al. Prevalence and etiology of community-acquired pneumonia in immunocompromised patients. *Clin Infect Dis.* 2019;68:1482–93.
- Fine MJ, Auble TE, Yealy DM, Hanusa BH, Weissfeld LA, Singer DE, et al. A prediction rule to identify low-risk patients with community-acquired pneumonia. *N Engl J Med.* 1997;336:243–50.
- Gilbert DN, Chambers HF, Saag MS, Pavia AT, Boucher HW, Black D, Freedman DO and Kim K. Schwartz. The Sanford guide to antimicrobial therapy, 52th, editor ed., 2022.
- Sollid JU, Furberg AS, Hanssen AM, Johannessen M. *Staphylococcus aureus*: determinants of human carriage. *Infect Genet Evol.* 2014;21:531–41.

22. ERS Task Force Report. Guidelines for management of adult community-acquired lower respiratory tract infections. *Eur Resp J*. 1998;11:986–91.
23. Lim WS, Baudouin SV, George RC, Hill AT, Jamieson C, Le Jeune I, et al. BTS guidelines for the management of community acquired pneumonia in adults: update 2009. *Thorax*. 2009;64(Suppl 3). iii1-55.
24. Corrêa Rde A, Lundgren FL, Pereira-Silva JL, Frare e Silva RL, Cardoso AP, Lemos AC, et al. Brazilian guidelines for community-acquired pneumonia in immunocompetent adults – 2009. *J Bras Pneumol*. 2009;35:574–601.
25. Corrêa RA, Costa AN, Lundgren F, Michelin L, Figueiredo MR, Holanda M, et al. 2018 recommendations for the management of community acquired pneumonia. *J Bras Pneumol*. 2018;44:405–23.
26. Infections IoMUFoE. The resistance phenomenon in microbes and infectious disease vectors: implications for human health and strategies for containment: workshop summary. 2003.
27. Ramsey C, Kumar A. H1N1: viral pneumonia as a cause of acute respiratory distress syndrome. *Curr Opin Crit Care*. 2011;17:64–71.
28. Lansbury L, Lim B, Baskaran V, Lim WS. Co-infections in people with COVID-19: a systematic review and meta-analysis. *J Infect*. 2020;81:266–75.
29. Youngs J, Wyncoll D, Hopkins P, Arnold A, Ball J, Bicanic T. Improving antibiotic stewardship in COVID-19: bacterial co-infection is less common than with influenza. *J Infect*. 2020;81: e55–e7.
30. Wee LE, Ko KKK, Ho WQ, Kwek GTC, Tan TT, Wijaya L. Community-acquired viral respiratory infections amongst hospitalized inpatients during a COVID-19 outbreak in Singapore: co-infection and clinical outcomes. *J Clin Virol*. 2020;128:104436.